

Interspecific Genetic Divergence in Sciaenids from Japan and Its Adjacent Waters

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Abstract Genetic divergence and phylogenetic relationships among *Nibea mitsukurii*, *Nibea albiflora*, *Pennahia argentata*, *Argyrosomus japonicus*, *Atrobuca nibe* and *Larimichthys crocea* were investigated by examining the electrophoretic patterns of 14 enzymes. The allele frequencies of 21 loci were estimated to calculate the genetic distances (D). The average D value among the sciaenid species increased in proportion to the level of taxonomic category. Relationships estimated by genetic markers well accorded with those estimated by morphological characters. The 6 sciaenid species were clearly divided into two distinct groups at a D value of 1.41.

Electrophoretic analysis of protein (especially enzyme) variants has been widely used in the study of many aspects of intraspecific- and species-level taxonomy. The reviews of Allendorf and Utter (1979), Avise (1975) and Ayala (1983) have shown that conspecific populations consistently demonstrate high levels of genetic (allozyme) similarity, as measured by Rogers (1972) or Nei's (1972) coefficients of genetic similarity, and that closely related species show much lower levels of genetic similarity. Related populations and species can thus be analysed for percentages of shared allozymes and these values used in making taxonomic decisions. Taniguchi et al. (1972, 1986) applied several biochemical markers detected by an electrophoretic method to the systematic study of fishes in the families Platycephalidae and Sparidae. These studies confirmed the principle that isozyme genes are convenient markers to estimate the

phylogenetic relationships of fishes.

This study utilizes isozyme gene markers to determine the genetic divergence and phylogenetic relationships among 6 sciaenid species (Pisces, Sciaenidae).

Materials and methods

Species names, number of individuals of each species tested, and sampling areas are given in Table 1.

The specimens were placed on dry ice upon capture and kept frozen until tested. The skeletal muscle, drumming muscle, liver, heart and eye were dissected from each individual specimen. The cell-lysate obtained by freezing and thawing was directly subjected to electrophoresis for phenotypic analysis. Starch gel electrophoresis was carried out following the procedure described by

Table 1. Scientific names (Trewavas, 1977), sampling locations and number of samples examined.

| Subfamily | Tribe | Species | Japanese name | No. of samples | Location |
|-------------|---------------|------------------------------|---------------|----------------|-----------------------------------|
| Otolithinae | Otolithini | <i>Pennahia argentata</i> | shiroguchi | 30 | Inland Sea, off Takamatsu |
| | | <i>Argyrosomus japonicus</i> | ohnibe | 6 | Tosa Bay, off Kochi |
| | | <i>Atrobuca nibe</i> | kuroguchi | 40 | East China sea, 32°15'N, 124°25'E |
| | Nibeini | <i>Nibea mitsukurii</i> | nibe | 40 | Tosa Bay, off Kochi |
| | | <i>Nibea albiflora</i> | koichi | 35 | Inland Sea, off Takamatsu |
| | Collichthyini | <i>Larimichthys crocea</i> | fuhsei | 7 | East China sea 32°15'N, 123°25'E |

Taniguchi and Numachi (1978), using a citric acid-aminopropylmorpholine (C-APM) buffer system of pH 6.0 (Taniguchi et al., 1978).

The list of enzymes, tissue specificity and loci are given in Table 2. A locus was considered polymorphic when the frequency of the most common allele was less than or equal to 0.99. When multiple loci coded for an enzyme, the locus with the most anodal migration was designated one, the next two, and so on. Allelic variants were designated according to their relative mobility, the most common allele being designated 100 and other alleles being given numbers that indicated their mobility relative to that of the common allele. Cathodal systems were designated in a similar way but were given a negative sign. The identity or difference of alleles at the same locus was decided by the banding position of isozymes on the same gel, based on the assumption that the isozyme bands migrating to the same position were composed of the same amino acid coded by the same gene. Genetic distance was calculated from the formula proposed by Nei (1972). The dendrogram was drawn by the unweighted pair-group method (UPGMA) (Sneath and Sokal, 1973).

Results and discussion

The electropherograms of isozyme were compared among the sciaenid species. The patterns clearly showed similarities and differences among the genera in the positions of the homopolymeric band for each isozyme locus. The electrophoretic pattern of fourteen enzymes enabled the identification of 21 separate loci (Table 2). These isozymes, obtained stably and clearly in all 6 species, were used as genetic markers. Table 3 shows the alleles and allele frequencies at each of the 21 loci for the 6 sciaenid species. Two to ten alleles were detected for each locus, and many loci were monomorphic and fixed in an allele. When the allele frequencies are compared between every two species, more number of divergent loci were found in more distantly related species pair in morphological characters (Taniguchi, 1969a, b, 1970).

In order to estimate the degrees of genetic divergence among the 6 species, the genetic distance (D) was calculated between every pair of species using the allele frequencies shown in Table 3. The D value will become zero when the allele frequencies of the two species compared are

Table 2. List of enzymes examined, loci identified and tissue assayed.

| Enzyme | Locus | Tissue assayed |
|---|--------------------------------|----------------------------------|
| Alcohol dehydrogenase (ADH) | <i>Adh</i> | liver |
| Aspartate aminotransferase (AAT) | <i>Aat-1</i> | liver |
| | <i>Aat-2</i> | liver, drumming muscle |
| Isocitrate dehydrogenase (IDH) | <i>Idh-1</i> | liver |
| | <i>Idh-2</i> | heart, drumming muscle |
| Glucosephosphate isomerase (GPI) | <i>Gpi-1</i> | drumming muscle |
| | <i>Gpi-2</i> | drumming muscle |
| Lactate dehydrogenase (LDH) | <i>Ldh-1</i> | eye |
| | <i>Ldh-2</i> | heart |
| | <i>Ldh-3</i> | muscle |
| 6-Phosphogluconate dehydrogenase (6-PGD) | <i>6-Pgd</i> | liver |
| Mannose phosphate isomerase (MPI) | <i>Mpi</i> | liver, heart |
| Esterase (EST) | <i>Est</i> | drumming muscle, eye, heart |
| Malate dehydrogenase (MDH) | <i>Mdh-1</i> | skeletal muscle, drumming muscle |
| | <i>Mdh-2</i> | heart |
| | <i>Mdh-3</i> | drumming muscle, skeletal muscle |
| Phosphoglucomutase (PGM) | <i>Pgm</i> | skeletal muscle |
| Super oxide dismutase (SOD) | <i>Sod</i> | liver |
| Fumarate hydratase (FM) | <i>Fm</i> | liver |
| Creatine kinase (CK) | <i>Ck</i> | drumming muscle |
| α -Glycerophosphate dehydrogenase (α -GPD) | <i>α-Gpd</i> | skeletal muscle |

Table 3. Allele frequencies at 21 loci in six sciaenid species. The numerals in parentheses show numbers of samples examined. Nm, *Nibea mitsukurii*; Na, *Nibea albiflora*; Aj, *Argyrosomus japonicus*; Pa, *Pennahia argentata*; An, *Atrobucca nibe*; Lc, *Larimichthys crocea*.

| Locus | Allele | Species | | | | | |
|--------------|--------|---------|--------|-------|--------|--------|-------|
| | | Nm(40) | Na(35) | Aj(6) | Pa(30) | An(40) | Lc(7) |
| <i>Aat-1</i> | 136 | 0.000 | 0.000 | 0.000 | 0.000 | 0.975 | 0.000 |
| | 120 | 0.000 | 0.014 | 0.000 | 0.000 | 0.025 | 0.000 |
| | 112 | 0.000 | 0.000 | 0.000 | 1.000 | 0.000 | 0.000 |
| | 105 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 0.786 |
| | 100 | 0.987 | 0.929 | 0.000 | 0.000 | 0.000 | 0.000 |
| | 84 | 0.000 | 0.000 | 1.000 | 0.000 | 0.000 | 0.000 |
| | 76 | 0.013 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 |
| | 72 | 0.000 | 0.057 | 0.000 | 0.000 | 0.000 | 0.214 |
| <i>Aat-2</i> | -50 | 0.038 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 |
| | -83 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 1.000 |
| | -100 | 0.962 | 1.000 | 1.000 | 1.000 | 1.000 | 0.000 |
| <i>Idh-1</i> | 119 | 0.000 | 0.014 | 0.000 | 0.000 | 0.000 | 0.000 |
| | 100 | 1.000 | 0.986 | 1.000 | 0.000 | 0.000 | 0.000 |
| | 92 | 0.000 | 0.000 | 0.000 | 0.000 | 1.000 | 0.000 |
| | 71 | 0.000 | 0.000 | 0.000 | 1.000 | 0.000 | 0.000 |
| | 56 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 1.000 |
| <i>Idh-2</i> | 212 | 0.000 | 0.029 | 0.000 | 0.000 | 0.000 | 0.000 |
| | 100 | 0.288 | 0.971 | 1.000 | 1.000 | 0.963 | 1.000 |
| | 25 | 0.712 | 0.000 | 0.000 | 0.000 | 0.038 | 0.000 |
| <i>Gpi-1</i> | 100 | 0.000 | 0.000 | 0.000 | 0.000 | 1.000 | 0.000 |
| | 93 | 0.000 | 0.000 | 0.000 | 0.897 | 0.000 | 0.929 |
| | 86 | 0.825 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 |
| | 83 | 0.000 | 0.000 | 1.000 | 0.000 | 0.000 | 0.000 |
| | 79 | 0.000 | 0.043 | 0.000 | 0.086 | 0.000 | 0.000 |
| | 66 | 0.175 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 |
| | 59 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 0.071 |
| | 55 | 0.000 | 0.957 | 0.000 | 0.017 | 0.000 | 0.000 |
| <i>Gpi-2</i> | -50 | 0.013 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 |
| | -100 | 0.986 | 0.986 | 1.000 | 0.983 | 0.863 | 1.000 |
| | -127 | 0.000 | 0.014 | 0.000 | 0.000 | 0.138 | 0.000 |
| | -130 | 0.000 | 0.000 | 0.000 | 0.017 | 0.000 | 0.000 |
| <i>Ldh-1</i> | 100 | 1.000 | 1.000 | 0.000 | 1.000 | 1.000 | 1.000 |
| | 50 | 0.000 | 0.000 | 1.000 | 0.000 | 0.000 | 0.000 |
| <i>Ldh-2</i> | 163 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 1.000 |
| | 100 | 1.000 | 1.000 | 1.000 | 1.000 | 1.000 | 0.000 |
| <i>Ldh-3</i> | 115 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 1.000 |
| | 106 | 0.000 | 0.000 | 1.000 | 0.000 | 1.000 | 0.000 |
| | 100 | 1.000 | 1.000 | 0.000 | 0.000 | 0.000 | 0.000 |
| | 97 | 0.000 | 0.000 | 0.000 | 1.000 | 0.000 | 0.000 |
| <i>Adh</i> | -24 | 0.000 | 0.000 | 1.000 | 0.000 | 0.000 | 0.000 |
| | -35 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 0.929 |
| | -55 | 0.000 | 0.000 | 0.000 | 0.000 | 1.000 | 0.000 |
| | -82 | 0.000 | 0.000 | 0.000 | 0.967 | 0.000 | 0.000 |
| | -88 | 0.000 | 0.071 | 0.000 | 0.000 | 0.000 | 0.000 |
| | -91 | 0.038 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 |
| | -94 | 0.000 | 0.000 | 0.000 | 0.033 | 0.000 | 0.071 |
| | -100 | 0.500 | 0.786 | 0.000 | 0.000 | 0.000 | 0.000 |
| | -112 | 0.000 | 0.143 | 0.000 | 0.000 | 0.000 | 0.000 |
| | -115 | 0.462 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 |

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Table 3. Continued.

| Locus | Allele | Species | | | | | |
|--------------------------------|--------|---------|--------|-------|--------|--------|-------|
| | | Nm(40) | Na(35) | Aj(6) | Pa(30) | An(40) | Lc(7) |
| <i>6-Pgd</i> | 112 | 0.000 | 0.000 | 1.000 | 0.000 | 0.000 | 0.000 |
| | 108 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 1.000 |
| | 100 | 0.787 | 1.000 | 0.000 | 0.000 | 0.000 | 0.000 |
| | 81 | 0.000 | 0.000 | 0.000 | 0.967 | 0.000 | 0.000 |
| | 77 | 0.213 | 0.000 | 0.000 | 0.000 | 0.986 | 0.000 |
| | 73 | 0.000 | 0.000 | 0.000 | 0.033 | 0.000 | 0.000 |
| | 58 | 0.000 | 0.000 | 0.000 | 0.000 | 0.014 | 0.000 |
| <i>Mpi</i> | 111 | 0.000 | 0.000 | 1.000 | 0.000 | 0.000 | 0.000 |
| | 108 | 0.000 | 0.000 | 0.000 | 0.000 | 0.225 | 1.000 |
| | 100 | 1.000 | 1.000 | 0.000 | 0.000 | 0.775 | 0.000 |
| | 96 | 0.000 | 0.000 | 0.000 | 0.183 | 0.000 | 0.000 |
| | 89 | 0.000 | 0.000 | 0.000 | 0.817 | 0.000 | 0.000 |
| <i>Est</i> | 112 | 0.000 | 0.000 | 0.000 | 0.000 | 1.000 | 0.000 |
| | 100 | 1.000 | 1.000 | 0.000 | 0.000 | 0.000 | 1.000 |
| | 97 | 0.000 | 0.000 | 1.000 | 0.000 | 0.000 | 0.000 |
| | 94 | 0.000 | 0.000 | 0.000 | 0.067 | 0.000 | 0.000 |
| | 88 | 0.000 | 0.000 | 0.000 | 0.933 | 0.000 | 0.000 |
| <i>Mdh-1</i> | 128 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 0.929 |
| | 117 | 0.000 | 0.000 | 0.000 | 1.000 | 0.000 | 0.000 |
| | 113 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 0.071 |
| | 100 | 1.000 | 1.000 | 0.000 | 0.000 | 0.013 | 0.000 |
| | 96 | 0.000 | 0.000 | 1.000 | 0.000 | 0.987 | 0.000 |
| <i>Mdh-2</i> | 100 | 1.000 | 1.000 | 0.000 | 1.000 | 0.000 | 0.000 |
| | 85 | 0.000 | 0.000 | 1.000 | 0.000 | 1.000 | 0.000 |
| | 60 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 1.000 |
| <i>Mdh-3</i> | -56 | 0.000 | 0.000 | 0.000 | 1.000 | 0.000 | 0.000 |
| | -100 | 1.000 | 1.000 | 0.000 | 0.000 | 0.000 | 0.000 |
| | -122 | 0.000 | 0.000 | 0.000 | 0.000 | 1.000 | 1.000 |
| | -178 | 0.000 | 0.000 | 1.000 | 0.000 | 0.000 | 0.000 |
| <i>Pgm</i> | 200 | 0.625 | 0.186 | 0.000 | 0.000 | 0.163 | 0.071 |
| | 100 | 0.375 | 0.814 | 1.000 | 0.967 | 0.837 | 0.929 |
| | -33 | 0.000 | 0.000 | 0.000 | 0.033 | 0.000 | 0.000 |
| <i>Sod</i> | 114 | 0.000 | 0.000 | 1.000 | 0.000 | 0.000 | 0.000 |
| | 103 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 1.000 |
| | 100 | 1.000 | 1.000 | 0.000 | 0.000 | 0.000 | 0.000 |
| | 76 | 0.000 | 0.000 | 0.000 | 1.000 | 0.000 | 0.000 |
| | 73 | 0.000 | 0.000 | 0.000 | 0.000 | 0.987 | 0.000 |
| | 35 | 0.000 | 0.000 | 0.000 | 0.000 | 0.013 | 0.000 |
| <i>Fm</i> | 100 | 0.000 | 0.000 | 1.000 | 0.917 | 0.986 | 1.000 |
| | 80 | 1.000 | 1.000 | 0.000 | 0.000 | 0.000 | 0.000 |
| | 68 | 0.000 | 0.000 | 0.000 | 0.083 | 0.014 | 0.000 |
| <i>Ck</i> | 157 | 0.000 | 0.000 | 1.000 | 0.000 | 0.000 | 1.000 |
| | 100 | 1.000 | 1.000 | 0.000 | 1.000 | 1.000 | 0.000 |
| <i>α-Gpd</i> | -33 | 0.000 | 0.000 | 0.000 | 0.000 | 0.987 | 0.000 |
| | -100 | 0.975 | 0.943 | 0.000 | 0.000 | 0.000 | 0.000 |
| | -115 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 1.000 |
| | -117 | 0.000 | 0.000 | 0.000 | 1.000 | 0.000 | 0.000 |
| | -150 | 0.000 | 0.000 | 0.000 | 0.000 | 0.013 | 0.000 |
| | -183 | 0.000 | 0.000 | 1.000 | 0.000 | 0.000 | 0.000 |
| | -233 | 0.025 | 0.057 | 0.000 | 0.000 | 0.000 | 0.000 |

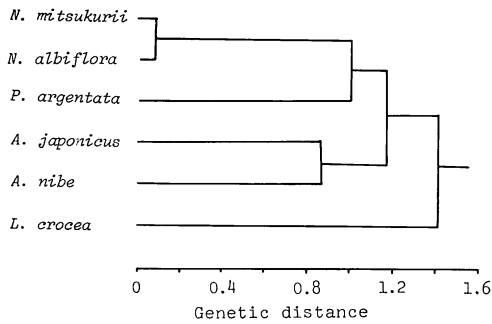


Fig. 1. Dendrogram showing the relationship among the 6 sciaenid species based on values of genetic distance.

completely identical, and will become infinite when the allele frequencies of the two species compared are completely divergent; namely, they do not share common alleles (Nei, 1972). Table 4 gives the list of genetic distances for each pair of species, and Table 5 shows the average genetic distance of

each pair of subpopulations or species for various levels of taxa. The average D value among species (*N. mitsukurii* and *N. albiflora*) belonging to the same genus was found to be 0.092, while that among genera belonging to the same subfamily was 1.212, ranging from 0.861 to 1.668.

To estimate the relationships between the 6 species, a dendrogram was drawn based on the genetic distances (Fig. 1). From this dendrogram, the six species were clearly divided into two major groups at a distance of 1.41. One group included *N. mitsukurii*, *N. albiflora*, *P. argentata*, *A. japonicus* and *A. nibe* while the other group consisted solely of *L. crocea*. The first group was further divisible into two groups at a distance of 1.2, one group consisting of *N. mitsukurii*, *N. albiflora* and *P. argentata*, and the other of *A. japonicus* and *A. nibe*. It was possible to further divide this first group into two subgroups at a distance of 1.0, one subgroup consisting of *N. mitsukurii* and *N. albiflora*, and the other of *P. argentata*.

Table 4. Estimates of genetic similarity and distance between 6 sciaenid species. Values above diagonal are estimates of genetic distance, those below are estimates of genetic similarity.

* The bias caused by small number of individuals was not corrected because the D values were larger than 0.15 (Nei, 1978).

| Species | 1 | 2 | 3 | 4 | 5 | 6 |
|-----------------------------------|-------|-------|-------|-------|-------|-------|
| 1. <i>Nibe mitsukurii</i> | | 0.092 | 1.460 | 1.080 | 1.085 | 1.668 |
| 2. <i>Nibe albiflora</i> | 0.912 | | 1.267 | 0.946 | 1.004 | 1.441 |
| 3. <i>Argyrosomus japonicus</i> * | 0.232 | 0.282 | | 1.251 | 0.861 | 1.428 |
| 4. <i>Pennahia argentata</i> | 0.339 | 0.388 | 0.286 | | 0.975 | 1.271 |
| 5. <i>Atrobucca nibe</i> | 0.338 | 0.366 | 0.423 | 0.377 | | 1.234 |
| 6. <i>Larimichthys crocea</i> * | 0.189 | 0.237 | 0.240 | 0.281 | 0.291 | |

Table 5. Average and range of genetic distance in sciaenid species.

| Item | Source of the data |
|-----------------------------------|---|
| Between conspecific subpopulation | $D=0.0017$ (0.00087–0.00220) (in <i>Pennahia argentata</i> from 4 locations based on 28 loci) $D=0.0390$ (in <i>Nibe albiflora</i> from 2 locations based on 29 loci) $D=0.0074$ (in <i>Nibe mitsukurii</i> from 2 locations based on 29 loci) |
| Between congeneric species | $D=0.0920$ (in genus <i>Nibe</i> on 29 loci) |
| Between consubfamilial genera | $D=0.3530$ $D=1.2120$ |

Genetic divergence between congeneric species has been observed in many fishes using biochemical markers. An average genetic distance between species has been reported as being 0.627 in 10 species of the genus *Lepomis* (Avisé and Smith, 1974a, b), 0.421 in 5 species of *Menidia* (Johnson, 1975), 0.214 in 2 species of *Moxostoma* (Buth, 1977), 0.229 in 3 species of *Campostoma* (Buth and Burr, 1978), 0.251 in 7 species of *Notropis* (Buth, 1979), 0.243 in 3 species of *Hypentelium* (Buth, 1980) and 0.353 in 3 species of the sciaenid genus *Cynoscion* (Fitzsimons et al., 1985). In the family Pleuronectidae, the average genetic distance was reported as being 0.01 between populations, 0.13 between subspecies, 0.62 between species and 1.11 between genera (Ward and Gelleguillos, 1983). In the family Carangidae, Kijima et al. (1986) indicated the average genetic distance as being 0.880 between species, 2.046 between genera and 2.290 between subfamilies, while in sparid fish Taniguchi et al. (1986) reported the average genetic distance as being 0.002 between conspecific subpopulations, 0.115 between congeneric species, 0.842 between consubfamilial genera and 1.273 between subfamilies. The average value found between consubfamilial genera in the sparid fish was almost identical to that found at the species level in carangid fish (Taniguchi et al., 1986). In fishes, as in other organisms (Ayala, 1983) the average genetic distance tends to be larger in higher taxa. This was reflected in the present study, where the *D* value between consubfamilial genera was larger than the *D* value between congeneric species, and that between congeneric species was larger than that between conspecific populations. The two species of *Nibea* (*N. mitsukurii* and *N. albiflora*) were, however, less diversified than the species level of the other genus, *Cynoscion* (Fitzsimons et al., 1985). The genetic distance was more or less equal to the subspecies level in published data (Taniguchi et al. 1986).

Taniguchi (1969a, b, 1970) studied the systematic relationships among the sciaenid species of the Japan and China Seas based on the osteology of the neurocranium, vertebrae, premaxillary and dentary. The present data support the classification based on the neurocranium. The sciaenid species in the present study were clearly divisible by the biochemical genetic markers into two distinct groups. One of the group included only *L. crocea*, while the other group, consisting of the

remaining 5 species, could be further divided at lower *D* levels into the *Nibea*-form (*N. mitsukurii* and *N. albiflora*), the *Pennahia*-form (*P. argentata*) and the *Argyrosomus*-form (*Argyrosomus* and *Atroubucca*). The *Pennahia*-form was found to be closer to the *Nibea*-form, while the *Argyrosomus*-form was intermediate between the *Pennahia*-form and *Pseudosciaena*-form (*Larimichthys*-form). Evidence obtained from the biochemical genetic study suggests that the *Pseudosciaena*-form evolved independently, and supports the result obtained from morphological studies.

The genetic distances thus give supplemental but important evidence pertaining to taxonomic problems on the recognition of species or subspecies which cannot be solved by conventional methods. It may be concluded that isozymes are useful markers to estimate relationships among species and to determine their taxonomic rank, as reported in earlier studies.

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日本およびその近海産ニベ科魚類の遺伝的分化

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日本産ニベ科魚類、ニベ、コイチ、シログチ、オオニベ、クログチおよびフーセイの6種について、14種類のアイソザイムの電気泳動像を検出し、それらの間の遺伝的分化と系統関係を調べた。21遺伝子座の対立遺伝子頻度を推定し、各魚種間の遺伝的距離(D)を求めた。D値は比較する2種間の分類学的位置関係が遠くなるほど大きくなった。遺伝標識にもとづいて推定された類縁関係は内部および外部形態にもとづく結果とよく一致した。ニベ科魚類はD値が1.41のところ、大きく2群に分けられた。

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